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The Absence of Significant Short-Term Electromagnetic Bioeffects in Giant Algal Cells Exposed to CW and Pulse-Modulated X-Band Bursts

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Abstract—Giant cells of the algae *Chara brauni* and *Nitella flexilis* were exposed to continuous wave and pulse-modulated bursts of X-band microwaves and the vacuolar potential was monitored for immediate radiation-correlated offsets. No such offsets were observed despite a resolution of approximately 5×10^5 , and despite the wide variety of frequencies, power levels, and pulse protocols employed.

I. INTRODUCTION

THE BIOLOGICAL EFFECTS of nonionizing electromagnetic radiation now appear to be numerous and

are being studied intensively (e.g., [1]–[4]). Those which seem to arise from direct electromagnetic heating of the test preparation (i.e., *thermal* effects) are generally the best characterized and least controversial. They tend to be associated with incident fields on the order of 100 W/m^2 (10 mW/cm^2) or greater. There are, however, in addition to putatively thermal effects, many others which, because of their occurrence at low power levels or because of the form of their variation with system parameters, can not readily be explained in ordinary thermal terms. These so-called *athermal* effects tend to be less well characterized and more controversial. They have been reviewed comprehensively by Adey [5].

Two characteristics common to most thermal and athermal phenomena are 1) a prolonged period of irradiation (at least minutes) is necessary for the effect to manifest itself and 2) the ability to resolve shifts in the vari-

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ables observed is small (less than 1 part in 10^2). In this laboratory, a divergent strategy has been adopted in which effects that occur early in the irradiation (within a few tens of milliseconds) are sought with very high resolution (greater than 1 part in 10^3) [6].

This paper presents an extension of these techniques into the 8.2–12.4-GHz (X-band) frequency range in an effort simply to detect the short term effects (if any) of low-level microwave irradiation.

II. MATERIALS AND METHODS

A. Materials

All experiments were conducted on the eukaryotic green algae *Chara braunii* and *Nitella flexilis* using the giant (roughly 10 mm long and 0.3 mm in diameter) internodal cells. Cells from *Chara* cultures were normally in the electrogenic state (i.e., vacuolar resting potential more negative than -120 mV) while those from *Nitella* cultures were in the nonelectrogenic state (i.e., vacuolar resting potential less negative than -95 mV) [7].

B. Electronic Methods

These were modified but slightly from those employed previously [6], [8].

The exposure apparatus consisted of a 1.0-mm-wide microstrip fabricated on a 0.62-mm-thick sheet of ceramic whose dielectric constant was 75. The line was terminated by a microwave chip resistor to match its approximately $12.5\text{-}\Omega$ characteristic impedance. A 2.2-mm-wide gap, perpendicular to the microstrip, formed a channel in the dielectric sheet down which an electrolyte solution at 25°C was flowed and into which the cell under test was placed with its upstream end under the microstrip. Because of the relatively good match 1) between cell and channel water, and 2) between channel water and dielectric sheet, the field should be uniform along the microstrip and the field pattern impressed upon the cell's upstream end should be that of the line's quasi-TEM mode.

The vacuolar resting potential of the cell was sensed by a glass micropipette inserted into the downstream end of the cell, and this potential was amplified and filtered as described previously [6]. Segments of the potential signal 400 ms long and phase locked to irradiation bursts were digitized and averaged for analysis. The average of 20 segments permitted a search for offsets on approximately the $3\text{-}\mu\text{V}$ level, a resolution of roughly 5 in 10^5 when compared to the vacuolar potential.

C. Irradiation Protocol

Bursts of irradiation 100 ms long were delivered to the exposure apparatus by a directional coupler and a double stub tuner. Nominal total power incident upon the fluid-filled channel was derived 1) by calibrating, frequency by frequency, the power to the tuner in terms of coupler output, 2) setting the tuner for negligible back reflection, 3) applying a theoretical correction factor for loss on the microstrip, and 4) allocating a 1.0-dB loss to the tuner. Nominal power density to the exposed end of the cell was

TABLE I
PROTOCOL FOR PULSE-BURST IRRADIATION AT 9.09 GHz*

	Pulse Interval (μs)				
	2000	1000	500	200	100
Pulse Width (μs)	2^{\S}	1^{\S}	0.5^{\S}	0.2^{\S}	0.1^{\S}
	20	10	5	2	1
	200	100	50	20	10
	2000	1000	500	200	100

*For most pulse protocols, the cells were tested at both 100 and 1000 W/m^2 (power averaged over the 100-ms burst); those protocols marked with the symbol \S were tested only at 100 W/m^2 .

derived by dividing this power by the cross-sectional area of the microstrip (0.62 mm^2).

To study the effects of frequency, nine different irradiation frequencies (8.20, 8.64, 9.09, 9.58, 10.08, 10.62, 11.18, 11.78, and 12.40 GHz) were used. This logarithmic coverage of the X-band was chosen so that any frequency-resonant offset of $Q \leq 20$ would be observed. The nominal power density of each 100-ms burst was $100\text{ W}/\text{m}^2$, and three different burst protocols were used: 1) continuous wave (CW); 2) 1.0- μs pulses 1 ms apart; 3) 0.1- μs pulses 0.1 ms apart.

To study the effects of pulse repetition rate, a frequency of 9.09 GHz was chosen and 100-ms bursts of the pulse structures described in Table I were employed.

To study the effects of lower power density, a frequency of 9.09 GHz was chosen and 100-ms bursts were applied at nominal power densities of 100, 50, 20, 10, 5, 2, and 1 W/m^2 . At each power density, both 1) CW bursts and 2) bursts of 1.0- μs -wide pulses separated by 1.0 ms were employed.

III. RESULTS

Each irradiation condition described above was tested on three electrogenic cells from *Chara braunii* and three nonelectrogenic cells from *Nitella flexilis*. In no instance was a significant radiation-correlated offset of the resting potential observed.

Fig. 1 shows two sample chart recordings containing summed responses of *Chara braunii* (a) and *Nitella flexilis* (b) to twenty radiation bursts. The traces reveal only background noise.

IV. DISCUSSION

It is evident that the resting potential showed no significant response under the irradiation protocols employed. Since the resting potential is commonly believed to be a sensitive indicator of general cellular function and condition, and since sufficiently powerful bursts of X-band irradiation are known to have observable effects [9], one must examine a number of hypotheses as to why the protocols employed caused no observed offsets.

First, there may be no cellular effects of significance. This is possible, but, in view of the present controversy

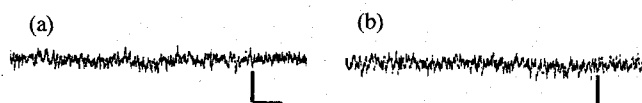


Fig. 1. (a) Electrogenic *Chara braunii* of resting potential -163 mV. Irradiation protocol: 100-ms bursts of 9.09 GHz radiation at an average nominal power density of 1000 W/m^2 ; each burst contained 200 5.0- μs pulses. (b) Nonelectrogenic *Nitella flexilis* of resting potential -60 mV. Irradiation protocol: 100-ms bursts of 11.18 GHz radiation at an average nominal power density of 100 W/m^2 ; each burst contained 1000 0.1- μs pulses. In each case, irradiation began about 60 ms after the beginning of the trace, vertical bar = $20 \mu\text{V}$, and horizontal bar = 40 ms.

about microwave bioeffects [10], such a supposition may well be unwarranted.

Second, there may be effects, but not on the vacuolar potential. Given the sensitivity of this variable to cellular conditions, this seems unlikely.

Third, there are X-band effects, but with such narrow frequency resonances that they were missed by the protocols used. Millimeter-wave effects of $Q \geq 1000$ are indeed known [11], [12], but such sharp resonances have not yet been reported in X-band.

Fourth, these protocols have effects but not ones which become manifest with the application of short, widely spaced bursts. This could well be the case. Unfortunately, the techniques employed here are less well suited to the study of long-term irradiation since the very high resolution achieved depended upon using the cell as its own control. This cannot be done over periods of many minutes because of slow millivolt-level stochastic fluctuation of the resting potential [13].

Thus, the principal importance of these data is that there appears to be no simple, rapid, obvious electrical effect of X-band irradiation in our preparation. The possibility of sharp resonances in power or frequency, or the existence of long-term effects, can not be ruled out. Moreover, these conclusions apply only to X-band and do not extend to higher (or lower) frequencies where there are theoretical reasons for expecting effects [14], [15].

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REFERENCES

- [1] M. A. Stuchly, "Interaction of radiofrequency and microwave radiation with living systems," *Radiat. Environm. Biophys.*, vol. 16, pp. 1-14, 1979.
- [2] K. Montaigne and W. F. Pickard, "Offset of the vacuolar potential of Characean cells in response to electromagnetic radiation over the range 250 Hz-250 kHz," *Bioelectromagn.*, vol. 5, pp. 31-38, 1984.
- [3] C. F. Blackman, J. A. Elder, C. M. Weil, S. G. Benane, D. C. Eichinger, and D. E. House, "Induction of calcium-ion efflux from brain tissue by radio-frequency radiation: Effect of modulation frequency and field strength," *Radio Sci.*, vol. 14, no. 65, pp. 93-98, 1979.
- [4] Y. H. Barsoum and W. G. Pickard, "Effects of electromagnetic radiation in the range 20-300 MHz on the vacuolar potential of Characean cells," *Bioelectromagn.*, vol. 3, pp. 193-201, 1982.
- [5] W. R. Adey, "Tissue interaction with nonionizing electromagnetic fields," *Physiol. Rev.*, vol. 61, pp. 453-514, 1981.
- [6] W. F. Pickard and Y. H. Barsoum, "Radiofrequency bioeffects at the membrane level: Separation of thermal and athermal contributions in the Characeae," *J. Membr. Biol.*, vol. 61, pp. 39-54, 1981.

- [7] N. A. Walker, "The transport systems of charophyte and chlorophyte giant algae and their integration into modes of behavior," in *Plant Membrane Transport: Current Conceptual Issues*, R. M. Spanswick, W. J. Lucas, and J. Dainty, Ed. Amsterdam: Elsevier, 1980, pp. 287-304.
- [8] Y. H. Barsoum and W. F. Pickard, "The vacuolar potential of Characean cells subjected to electromagnetic radiation in the range 200-8200 MHz," *Bioelectromagn.*, vol. 3, pp. 393-400, 1982.
- [9] A. V. Gokhale, K. Montaigne, and W. F. Pickard, "Vacuolar hyperpolarizing offsets in Characean cells exposed to mono- and bichromatic CW and to squarewave modulated electromagnetic radiation in the band 200-1000 MHz," Submitted to *Bioelectromagn.*, 1983.
- [10] "Experiments show microwaves can damage chromosomes," in *The Institute*, [News Supplement to *IEEE Spectrum*], vol. 7, p. 1, Aug. 1983.
- [11] W. Grundler and F. Keilmann, "Nonthermal effects of millimeter microwaves on yeast growth," *Z. Naturforsch.*, vol. 33, pp. 15-22, 1978.
- [12] W. Grundler, F. Keilmann, V. Putterlik, and D. Strube, "Resonant-like dependence of yeast growth rate on microwave frequencies," *Brit. J. Cancer.*, vol. 45, pp. 206-208, 1982.
- [13] R. L. Roa and W. F. Pickard, "The use of membrane electrical noise in the study of Characean electrophysiology," *J. Exp. Bot.*, vol. 27, pp. 460-472, 1976.
- [14] H. Fröhlich, "The biological effects of microwaves and related questions," *Adv. Electronics and Electron Phys.*, vol. 53, pp. 85-152, 1980.
- [15] W. F. Pickard and F. J. Rosenbaum, "Biological effects of microwaves at the membrane level: Two possible athermal electrophysiological mechanisms and a proposed experimental test," *Math. Biosci.*, vol. 39, pp. 235-253, 1978.



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